

Journal of Agriculture, Food, Environment and Animal Sciences Tarım, Gıda, Çevre ve Hayvancılık Bilimleri Dergisi http://www.jafeas.com, ISSN: 2757-5659 J. Agric. Food, Environ. Anim. Sci. 6(1): 31-49, 2025

# Leaf-Based Varietal Categorization of Sweetpotato (*Ipomoea batatas* L. Lam.), a Potentially Healthful Vegetable, Using Image Processing and K-Means Clustering

Shahidul ISLAM<sup>1\*</sup>, Towfiqur RAHMAN<sup>2</sup>, Md. Hamidul ISLAM<sup>1,3</sup>, Abdul MOMIN<sup>4</sup>

<sup>1</sup>Department of Agriculture, University of Arkansas at Pine Bluff, 1200 N. University Drive, AR 71601, USA <sup>2</sup>Department of Farm Power and Machinery, Sylhet Agricultural University, Sylhet-3100, BANGLADESH <sup>3</sup>Department of Farm Power and Machinery, Bangladesh Agricultural University, Mymensingh-2202, BANGLADESH

<sup>4</sup>Agricultural Engineering Technology, School of Agriculture, Tennessee Tech University, Cookeville, TN 38505, USA

<sup>1</sup>https://orcid.org/0000-0003-3131-6692, <sup>2</sup>https://orcid.org/0000-0001-9520-2753 <sup>3</sup>https://orcid.org/0000-0002-5115-091X, <sup>4</sup>https://orcid.org/0000-0002-3139-5274

\*Corresponding author: islams@uapb.edu

Research Art	icle ABSTRACT	
Article Histo Received:8 Au Accepted:14 Ja <u>Published onlit</u> Keywords: Sweetpotato Image Proces K-Means Clu Phenotype E Health Benef	gust 2024 muary 2025 he: 01 June 2025of phenolic compounds in health promotion. leaves varies from varied identifying factors for detecting sweetpotato systems. This study tsLeaves sing stering valuation tsdetecting sweetpotato systems. This study (RGB) of two sweetpot extract relevant feath characteristics, i.e., lead leaf ratio, between the algorithm identified two centroid values (Clust Results revealed that sp prominent physical of demonstrates the pro processing techniques The results bridge th assessment, fostering a	<i>atatas</i> Lam) leaves contain higher concentrations of, flavonoids, and carotenoids that are remarkable However, the nutrient content in sweetpotato ety to variety, and leaf shape and color are the key the varietal classification of sweetpotatoes. So, leaves is essential for the in-situ identification of and for developing intelligent agricultural aimed to create a leaf-shape-based varietal the for sweetpotato using image processing th a K-means clustering algorithm. 38 leaf images ato cultivars were collected and pre-processed to tres. A distinct difference in leaf physical area, perimeter, circularity factor, breadth, and two varieties was observed. K-means clustering to sweetpotato varieties as distinct clusters with er 0: Area 695627 and Cluster 1: Area 525895). weet potato leaves in cluster 0 tend to have more characteristics than in cluster 1. This result spects of using machine learning and image for in situ varietal classification of sweetpotato. e visual characteristics and their quantitative deeper understanding of the plant's phenotype ncements in agriculture, research, and crop
TaCita		25. Leaf-Based Varietal Categorization of Sweetpotato
To Cite :	Clustering. Agriculture, Food, Environment and	ful Vegetable, Using Image Processing and K-Means d Animal Sciences, 6(1): 30-49.

## INTRODUCTION

Sweetpotato (*Ipomoea batatas* L.) is often referred to as a superfood and plays a pivotal role in ensuring food security worldwide. It is a potent source of essential nutrients such as vitamins A and C, dietary fiber, and minerals. Anthocyanins and carotenoids are potent antioxidants and offer several health advantages, including lowering the risk of cancer and heart disease, assisting blood sugar regulation, and reducing obesity. They are abundant in sweet potatoes (Islam, 2006; 2024). Sweetpotato is gaining popularity in the United States as a very healthy vegetable. The per capita intake of the sweetpotato increased by about 56% from 2000 to 2014 (Statista, 2016). Recent research on nutrients and phenolics has encouraged agriculturists and food scientists to investigate the beneficial characteristics of sweet potato leaves for human health. Several studies have reported that sweetpotato leaves offer remarkable health benefits, making them a valuable addition to any diet (Islam, 2008; Xu et al., 2010; Taira et al., 2013; Wang et al., 2016).

The increasing awareness of the health benefits of sweetpotatoes and their reputation as an alternative vegetable crop has been noticed by the food processors who have developed products that include sweetpotato fries, puffs, wedges, crisps, sticks, pie, and mashed sweetpotatoes that are now available in many local stores (Islam, 2006; Smith et al., 2009). Sweetpotato leaves contain high concentrations of polyphenolics compared with the major commercial vegetables such as spinach, broccoli, cabbage, and lettuce. Extensive reports have revealed that sweetpotato leaves can protect the human body from oxidative damage, inflammation, aging, and hypertension due to various antioxidant compounds, including phenolic compounds, flavonoids, vitamin C, and carotenoids (Gunathilake and Ranaweera, 2016; Rumbaoa et al., 2009). Several researchers claimed that sweetpotato leaves are superior in health-associated functionality compared to petioles, stems, tuberous roots, and many other commercial leafy vegetables (Lako et al., 2007; Xu et al., 2010; Huang et al., 2013).

Sweetpotato has a wide variability in phenotypic and botanical characteristics and is readily differentiated based on the shape of leaves and other agronomic characteristics (Zhang et al., 2000; Amankwaah, 2012). Sweetpotato leaves are medium to large and are cordate or heart-shaped with pointed tips. The leaves grow in an alternate pattern and may be palmate. Sweet potato leaves range in color from dark to yellow-green or purple and tend to be darker on the surface and lighter on the underside. According to the published report, the sweetpotato variety substantially affected antioxidant levels and the qualities of buds, leaves, petioles, and stems (Jia et al., 2022). Variety with pure green leaves had higher total phenolic content and antioxidant activity in buds, leaves, petioles, and stems. Still, the variety with green-purple leaves contained substantially more anthocyanins than green and yellow leaves (Chen et al., 2018; Jia et al., 2022). Also, the variation in nutritional quality among different sweetpotato

varieties significantly impacts nutritional adequacy. Furthermore, other factors such as variety, environmental circumstances, and disease susceptibility influence sweetpotato plant productivity and quality (Ngailo et al., 2013). Therefore, sweet potato varietal categorization based on morphological features such as leaf size, shape, color, and texture are required for diversity evaluations for both plant genetic resource conservation and usage.

Conventional methods for the measurement and analysis of morphological characteristics in plant breeding are labor-intensive, costly, and time-consuming and increase the chance of generating errors in trait measurements (Kumar et al., 2015). However, the demand for rapid progress in genetic gain within breeding programs has led to a growing emphasis on the development and adoption of reliable, automatic, multifunctional, and high-throughput phenotyping technologies (Zhao et al., 2019). These technologies facilitate the rapid and accurate collection of vast amounts of morphological data, enabling more efficient and precise breeding strategies. Recent advancements in image analysis have revolutionized morphological characterization, offering a nondestructive and time-efficient alternative to conventional methods (Gehan et al., 2017). By automating the process, researchers can collect vast amounts of morphological data quickly and accurately, leading to more efficient breeding programs. Image-based phenotyping, in particular, has gained significant traction due to its ability to integrate computer vision, pattern recognition, and data mining approaches with advanced machine learning algorithms (Chitwood et al., 2013; Gehan et al., 2017). Several researchers have demonstrated the potential of combining these technologies to analyze plant morphology and phenotypic characteristics. For example, Du et al. (2006) utilized image processing techniques to extract morphological features and invariant moment features of various shapes of different plant leaves to classify leaf species. Lu et al. (2023) used image-based analysis integrated with machine learning to distinguish rice varieties based on panicle traits. Wang et al. (2023) developed a convolution block attention module and added to CNN framework to enhance and extract critical features of leaf images and results showed that the proposed leaf detection method outperforms state-of-the-art object detection methods. However, this study did not consider any classification approach for varietal classification of sweetpotato. This study focuses on the development of a leaf-shapebased varietal classification technique for sweetpotato using image processing techniques coupled with a K-means clustering algorithm. The study aims to improve sweet potato research and provide valuable insights into identifying and categorizing leaf variants. These findings have practical applications in agriculture, breeding programs, and crop management, supporting more efficient and informed decisionmaking processes.

# MATERIAL and METHOD

## Sample Collection

The experimental sweetpotato leaf samples were collected from UAPB Agricultural Experimental Farm in Pine Bluff, Arkansas. Two different genotypes, UAPB 18 (Variety 1) and UAPB 39 (Variety 2), were selected from thirty-eight accessions of sweetpotato leaves initially used. Each of the nineteen images was used in this study. After collecting the sweetpotato leaf samples, the leaves were cleaned to remove dirt and foreign matter from the leaf surface. This experiment was conducted at the Horticulture Laboratory at the University of Arkansas at Pine Bluff (UAPB), Arkansas, USA.

# Analysis of Vitamin C, Oxalate and Chlorophyll

The accessions' Vitamin C, oxalate, and chlorophyll content were determined using the spectrophotometric method described previously (Islam et al., 2023; Alam et al., 2020; Li et al., 2017).

## **Extraction and Measurement of Total Anthocyanin**

The leaf samples (500 mg) had 10 mL of 0.5% H2SO4 solution added and were steeped overnight at room temperature. A color value (CV) for the extract, which is an indicator of total anthocyanin, was determined using the formula:  $CV=0.1 \times OD 530 \times 4 \times 20/g$  DW, where OD530 is a spectrophotometric reading (530 nm), four corrects for the dilution, and 20g DW represents the dry weight of the sample (Islam et al., 2005).

## **Image Acquisition**

Due to its availability, high performance, low price, and portability, the cellphone camera has emerged as a potential digital imaging device (Liang et al., 2005). Si et al. (2016) reported promising results in the case of potato tuber grading when they used images captured under normal room lighting conditions. Considering these recent endeavors, the images of sweet potato leaves used in this research were acquired using a 12 MP camera without flash under stationary conditions. A focal distance of about 25 cm was maintained during the image acquisition to obtain quality images. The image resolutions were 1536 x 2048 pixels each and were stored in jpeg format. Besides, an image of a measuring scale was acquired for further use as a reference for dimensional calibration. The acquired sweet potato leaf images were sampled and denoted as Variety 1 and Variety 2 for further analysis.

## **Image Processing**

The image processing operation includes image preprocessing, color space conversion, background segmentation, and feature extraction. Since acquired images may have

distortion and shadow effects (which could cause challenges with segmentation and feature extraction), image preprocessing was done to obtain a better-quality image (Li et al., 2017). The segmentation techniques for removing the background from an image include thresholding and clustering. This work used a color thresholding method to eliminate the background from each sweet potato image. The MATLAB Color Thresholder App within the MATLAB® computational environment was used for color thresholding (MathWorks, USA, version R2020b).

First, the RGB (red, green, and blue) image was converted to HSV (hue, saturation, value) color space, which defines the threshold value based on the channel histogram. Then, a masked image was created based on the chosen histogram threshold setting and applied with the *imfill* and *bwareaopen* operations to eliminate tiny (pixel <100) artifacts that may appear during the data collection (Yao et al., 2009; Islam et al., 2021). Finally, the masked output image was obtained for the segmented sweet potato leaf sample image (Figure 1). These procedures were repeated for all potato sample images. Then, the feature extraction algorithm was run to collect the minor axis, major axis, surface area, and perimeter for further classification model development.



Figure 1. Example of processed sweetpotato leaf images

Each selected leaf characteristic was subjected to the following measurements explained in

Table 1. The leaf characteristics data was then exported into a Microsoft CSV file for calculating the breadth ratio and circularity factor. The breadth ratio was calculated by dividing max length by breadth, and the circularity factor was calculated using Equation 1 (Sun et al., 2017).

Circularity Factor (CF) = 
$$4\pi \frac{A}{P^2}$$
 (Equation 1)

Where P is the perimeter of the leaf in pixels; A is the area of the leaf in pixels.

Parameter	Description
Area*/	The total area of the leaf, measured in pixels
Perimeter	The length of the leaf's outer boundary is measured in pixels.
Circularity Factor (CF)	A dimensionless number represents the roundness of the leaf shape. CF value closer to 1 indicates more circular shapes.
Maximum Length	The longest dimension of the leaf, measured in pixels.
Breadth	The maximum width of the leaf is perpendicular to the maximum length,
	measured in pixels.

Table 1. Explanation of leaf characteristics measurement

## **Statistical Analysis**

Descriptive Statistics (DS) such as mean, standard deviation, minimum, and maximum values were computed for each measured leaf characteristics parameter. Using two-sample Z-tests based on the DS value, statistically significant differences between the two sweetpotato varieties were examined.

## **K-means Clustering**

The K-means clustering algorithm is a popular method to solve clustering cases. The basic concept behind K-means clustering is to divide an input data set into a predetermined number of clusters (k) and calculate the starting centroid for each cluster (Sun et al., 2017). Here, the optimal number of clusters (k) from the sweetpotato leaf dataset was determined using the Elbow approach. The Elbow method iterates over a range of cluster numbers, computing the sum of squared distances (inertia) of samples to the nearest cluster center for each cluster number. The number of clusters is plotted against the relevant inertia values to determine the optimal cluster number (Figure 2). The value of k is chosen when the rate of decrease in inertia slows sufficiently, and the graph begins to resemble a straight line. Later, the predefined number of clusters (k = 2) generated from Elbow Analysis was utilized to perform K-means clustering with the Scikit-Learn library.

Islam et al., / J. Agric. Food, Environ. Anim. Sci. 6(1): 31-49, 2025



Figure 1. K-means clustering Elbow method to define the number of clusters

#### **RESULTS and DISCUSSION**

The sweetpotato has become a component of an ever-increasing range of products. Internationally, the plant has diverse uses, including ornamental, livestock feed, starch and alcohol manufacture, human consumption, biofuel, and bioplastic production. Recently, research has been conducted to determine the health-promoting functions of sweetpotato (**Table 2**). The following aspects of these functions are important when considering new uses for sweetpotato storage roots and leaves.

Physiological Function	Related components	References
Antioxidative activity/	Polyphenol, anthocyanin	(Islam, 2006; 2008; 2014; 2016; 2019;
Radical scavenging activity		2024)
(Leaves & tuber)		
Antimutagenicity	Polyphenol, anthocyanin	(Islam, 2006; 2008; 2019; Suda et al.,
(Leaves & tuber)		1998; Peluso et al., 1995)
Anticarcinogenesis	Polyphenol, anthocyanin	(Islam, 2006; 2009; 2019; Islam et
(Leaves & tuber)		al., 2020; Shimozono et al., 1996)
Antihypertension	Polyphenolics, anthocyanin	(Suda et al., 1998; Matsui et al.,
(Leaves & tuber)		2002; 2004)
Antimicrobial activity	Fiber, pectin-like	(Yamakawa and Yoshomoto, 2002;
(Leaves & tuber)	polysaccharide	Yoshomoto, 2001)
Antiinflammation	Polyphenol	(Matsui et al., 2002; 2004)
(tuber)		
Antidiabetic effect	Anthocyanin, polyphenol	(Toeller, 1994; Matsui et al., 2004)
(Leaves & tuber)		
Anti-HIV	Polyphenolics	(Mahmood et al., 1993)
(Leaves & tuber)		
Promotion of	Dietary fiber	(Yoshimoto, 2001; Yamakawa and
Bifidobacterium growth		Toshimoto, 2002)
(Leaves & tuber)		
Reduction of liver injury	Polyphenol	(Suda et al., 1998)
(tuber)		

Table 2. Beneficial function of sweetpotatoes

#### **Pigments and Nutritional Attributes**

The chlorophyll, vitamin C, oxalate, and total anthocyanin content of sweetpotato leaves is shown in Table 3. The variation of oxalate contents among the varieties was found to be significant. The oxalate content of sweetpotato leaves was lower than that of Amaranthus (Onyango et al., 2008; Radek and Savage, 2008; Ogunlesi et al., 2010).

Table 3. Chlorophyll, Anthocyanin, Vitamin C, and Oxalate contents of the sweet potato leaves were studied.

Accessions	Chlorophyll contents (mg/100g DW*)	Total Anthocyanin (color value/g DW)	Vitamin C (nmol ascorbate /10g DW	Oxalate (nmol oxalate /10g DW)
UAPB 18	512.18** ± 2.72	$22.29 \pm 1.17$	$39.59 \pm 2.11$	1761 ± 12.19
UAPB 39	$684.41 \pm 3.91$	$18.62 \pm 1.07$	$46.83 \pm 3.07$	2272 ± 15.32
*DIN D				

\*DW= Dry weight; \*\*n (number of observations) = 15

Moreover, some vegetables grown in India were reported to contain high oxalate ranging from 5138 to 12576 mg/100g DWB (Radek and Savage, 2008). The ascorbate content ranges from 39.59 to 56.83 nmol/10 mg DW. A significant variation of results among varieties was observed. Our results are comparable to those of several authors who determined vitamin C in various sweetpotato leaves (Onyango et al., 2008; Away et al., 2013). Significant variations between the leaf chlorophyll and anthocyanin contents of the two accessions studied were also found. The study examined the chlorophyll, anthocyanin, vitamin C, and oxalate contents of sweetpotato leaves. By analyzing these components, researchers aimed to assess the nutritional value and potential health benefits of sweetpotato leaves. Chlorophyll content was measured to understand the plants' photosynthetic efficiency and overall health. Anthocyanin levels were assessed for their antioxidant properties, while vitamin C content was evaluated due to its essential role in human nutrition. Additionally, oxalate levels were analyzed to determine their potential impact on health, particularly calcium absorption. The findings provide valuable insights into the nutritional profile of sweetpotato leaves, highlighting their potential as a healthful addition to the diet (Onyango et al., 2008; Radek and Savage, 2008; Ogunlesi et al., 2010; Away et al., 2013).

#### Leaf Characteristics Analysis

A statistical analysis was conducted to evaluate the significant differences in leaf characteristics between the two sweet potato varieties, as summarized in Table 4. The results revealed that the mean leaf area for variety 1 was 534,146 square pixels, while variety 2 exhibited a significantly larger mean area of 648,741 square pixels, suggesting that variety 2 has larger leaf surfaces, potentially enhancing its photosynthetic efficiency. In contrast, the mean leaf perimeter of variety 1 was significantly higher at 6,251 pixels, compared to 4,026 pixels for variety 2, indicating a more compact leaf shape in variety 2. The compactness factor (CF), which reflects the degree of leaf compactness, was also significantly higher in variety 2, reinforcing its compact morphology. Additionally, the maximum leaf length and breadth were both larger in variety 1, with a mean length of 1,286 pixels and breadth of 1,235 pixels, compared to 1,191 pixels and 942 pixels, respectively, for variety 2. Despite its smaller dimensions, variety 2 had a higher mean leaf ratio (1.28) than variety 1 (1.06), indicating that the leaves of variety 2 are more elongated relative to their width. These findings highlight distinct morphological differences, with variety 2 producing larger, more compact, and elongated leaves, while variety 1 exhibits broader leaves with larger perimeters. Such differences may have implications for the varieties' adaptability, growth efficiency, and suitability for specific agricultural practices.

Variety	Parameters	Leaf area	Perimeter	CF	Max Length	Breadth	Ratio
	Min	396847	4892	0.110	994	872	0.730
	1Q	474116	5908	0.147	1245	1157	1.000
1	Med	542463	6219	0.170	1302	1234	1.040
1	Mean	534146	6251	0.175	1286	1235	1.059
	3Q	593288	6699	0.192	1350	1358	1.070
	Max	694882	8096	0.260	1486	1567	1.590
	Min	503626	3485	0.360	884	732.0	0.800
	1Q	560862	3912	0.450	1143	926.0	1.222
0	Med	662966	4006	0.500	1212	956.0	1.275
2	Mean	648741	4026	0.5039	1191	941.6	1.278
	3Q	724265	4232	0.5625	1263	990.5	1.320
	Max	867348	4424	0.630	1409	1108.0	1.690

Table 4. Statistical summary of sweetpotato leaf phenotypic characteristics

Table 5. Two-sample Z-test results

Parameter	Z-Score	P-Value	Significant $(\alpha=0.05)$
Leaf area	-4.19	0.000028	Yes
Perimeter	19.28	0.000000	Yes
CF	-20.07	0.000000	Yes
Max length	4.41	0.000011	Yes

Islam et al., / J. Agric. Food, Environ. Anim. Sci. 6(1): 31-49, 2025

Breadth	10.28	0.000000	Yes
Ratio	-13.45	0.000000	Yes

The two sample Z-test results (Table 5) revealed significant differences between the two sweet potato varieties across all measured leaf parameters at a significance level of  $\alpha$ =0.05. Variety 2 exhibited significantly higher mean values for Leaf Area and Compactness Factor (CF), as indicated by negative Z-statistics. In contrast, Variety 1 showed significantly higher mean values for Perimeter, Max Length, Breadth, and Ratio, with positive Z-statistics. The extremely low p-values (p<0.0001, p< 0.0001, p<0.0001) for all parameters confirm the robustness of these differences, suggesting that the two varieties have distinct morphological characteristics in their leaves. These findings highlight the potential variability in traits between the varieties, which could have implications for their growth, productivity, and adaptability.

#### **K-means** Clustering

The K-means clustering algorithm was applied to the dataset to explore the clustering patterns and potential groupings within the sweet potato leaf samples and evaluate the results with statistical inferences. The analysis revealed two distinct clusters corresponding to the two sweet potato varieties (varieties one and two). The results indicate that one cluster had 53% of the total samples, while another had 47%.

Table 6. Centroid position for characteristics of two different sweet potato leaf varieties

Cluster	Perimeter	Area	CF	Max Length	Breadth
0	4539	695627	0.481	1247	1018
1	5580	525895	0.243	1237	1141

Table 6 summarizes the centroid values for key morphological characteristics of two distinct clusters of sweet potato leaf varieties. Cluster 0 is characterized by a perimeter of 4539 units and a larger leaf area of 695,627 units, indicating broader leaf structures compared to Cluster 1, which has a perimeter of 5580 units and a smaller area of 525,895 units. The Compactness Factor (CF), which reflects the shape compactness, is higher in Cluster 0 (0.481), suggesting more compact leaf shapes, while Cluster 1 has a lower CF (0.243), indicating more elongated or irregular shapes. Both clusters exhibit similar maximum lengths, with Cluster 0 at 1247 units and Cluster 1 at 1237 units. However, the breadth of Cluster 0 is slightly narrower (1018 units) than that of Cluster 1 (1141 units). These morphological differences highlight the structural diversity between the two sweetpotato leaf varieties.

A scatter plot of area versus perimeter (Figure 3) was used to visualize the clusters, providing insights that the leaf area and the perimeter have contributed to the most separation of these varieties. K-means clustering analysis on sweet potato leaf samples used leaf area, perimeter, CF, max length, and breadth features. The clustering analysis

resulted in two distinct clusters with well-defined centroid values. In Table 4, the centroid of cluster 0 had higher values for features like area, CF, and max length than cluster 1. This result suggests that sweet potato leaves in cluster 0 tend to have more prominent physical characteristics of sweetpotato leaves. Figure 4 shows the clear separation of two varieties of sweetpotato leaves.



Figure 3. K-means clustering of sweet potato samples based on leaf area and perimeter Note: The values are represented in pixels; the x mark represents the centroid of each variety.

#### **Identification Important Features**

Sweetpotato leaves exhibit distinct phenotypic characteristics that contribute to their identification and classification. The leaves are typically heart-shaped or lobed, with various variations in leaf margins, including entire, serrated, or deeply lobed edges. The leaf color varies, encompassing shades of green, and in some varieties, a purple pigmentation may be present (Away et al., 2013). The leaves are arranged alternately along the stem, and their size can vary significantly among different cultivars. Sweetpotato leaves often display prominent venation, and the leaf surface may be smooth or textured. Leaf structure plays a crucial role in photosynthesis, and the plant's adaptive characteristics are reflected in these phenotypic traits. Understanding the diverse phenotypic characteristics of sweetpotato leaves is essential for botanical classification and agricultural practices related to cultivation and breeding (Jackson et al., 2020). Image analysis of sweetpotato leaves involves various techniques to study and interpret visual data captured from the leaves. Common aspects analyzed include leaf morphology, color, texture, and signs of diseases or stress. Digital imaging tools, such as computer vision algorithms, can be applied to quantify features like leaf size, shape, and pigmentation. Chlorophyll content, an indicator of plant health, can be assessed through color analysis. Texture analysis may reveal patterns associated with specific conditions or diseases affecting the leaves. Additionally, machine learning

models can be trained on image datasets to automate the identification of different leaf characteristics or potential issues (Gupta et al., 2020). Overall, image analysis in sweetpotato leaves aids researchers and farmers in understanding plant health, optimizing cultivation practices, and detecting any abnormalities for timely intervention. The relationship between image analysis and phenotypic characteristics of sweetpotato leaves is symbiotic, as image analysis is a powerful tool to quantify and interpret these observable traits. Image analysis allows for the objective and precise measurement of various phenotypic features, enhancing understanding of sweetpotato leaf morphology and health. For instance, size, shape, color, and textureintegral components of phenotypic characteristics - can be quantified using computer vision algorithms (Gupta et al., 2020; Su and Xue, 2021). Using image analysis, researchers can systematically evaluate and compare sweetpotato varieties based on their leaf attributes. This quantitative data aids in identifying patterns, assessing genetic diversity, and facilitating breeding programs to enhance desirable traits. Moreover, image analysis provides a non-invasive means of monitoring plant health, enabling the early detection of stress, diseases, or nutrient deficiencies by analyzing changes in leaf color, texture, or other visual indicators (Rosero et al., 2019; Su and Xue, 2021; Wang et al., 2023).

Image analysis of sweetpotato leaves finds diverse applications across agriculture, research, and crop management (Rosero et al., 2019; Jackson et al., 2020; Gupta et al., 2020; Su and Xue, 2021; Wang et al., 2023). Here are several critical applications: (i) Phenotypic Characterization: Image analysis enables the quantitative assessment of phenotypic characteristics such as leaf size, shape, color, and texture. This aids in cataloging and comparing different sweetpotato varieties, supporting breeding programs to develop crops with desired traits. (ii) Disease Detection and Monitoring: Early detection of diseases is crucial for crop management. Image analysis helps identify visual cues associated with diseases, such as discoloration or pattern changes in sweetpotato leaves, allowing for timely intervention and preventing the spread of infections, (iii) Nutrient Deficiency Identification: Leaf color and morphology changes can indicate nutrient deficiencies. Image analysis assists in recognizing these visual cues, helping farmers adjust fertilization practices to optimize nutrient levels for healthy sweetpotato growth. (iv) Stress Assessment: Sweetpotato plants respond to various environmental stresses. Image analysis allows for quantifying stress-related changes in leaves, such as wilting or discoloration, helping researchers and farmers assess the impact of environmental factors on plant health. (v) Yield Prediction: By analyzing leaf characteristics throughout the growing season, image analysis contributes to predicting sweetpotato yield. This information is valuable for farmers planning harvest schedules and optimizing crop production, (vi) Precision Agriculture: Image analysis supports precision agriculture practices by providing detailed spatial information about the crop. This includes leaf characteristics variations across different field areas, allowing for targeted interventions and resource optimization. (vii) Research and Development: Image analysis is fundamental in research endeavors, facilitating the study of sweetpotato genetics, physiology, and responses to various conditions. It expedites data collection and analysis, aiding scientists in making informed decisions and advancing knowledge in sweetpotato cultivation. Therefore, image analysis of sweetpotato leaves is a versatile tool with applications ranging from crop improvement to disease management, offering valuable insights for sustainable and efficient sweetpotato cultivation (Rosero et al., 2019; Jackson et al., 2020; Gupta et al., 2020; Su and Xue, 2021; Wang et al., 2023).

## CONCLUSION

In conclusion, there has been an increase in the production and consumption of sweet potato and its leaves. This study demonstrates how image processing techniques and clustering algorithms can be used to analyze sweet potato leaves, providing valuable insights into the different morphological characteristics of two sweet potato varieties. These insights can help improve crops, develop disease-resistant breeding programs, and manage agricultural systems more intelligently. The research focuses on categorizing sweetpotato varieties based on their leaf characteristics, leveraging advanced image processing techniques and K-means clustering for accurate identification and classification. , This study aims to enhance the precision of varietal identification using computer vision system based on leaf features like shape, color, and texture. Results revealed that distinct morphological differences exist between two varieties, with variety 2 producing larger, more compact, and elongated leaves, while variety 1 exhibits broader leaves with larger perimeters. This approach benefits the agricultural management and breeding of sweet potatoes and promotes the inclusion of this nutritious vegetable in our diets. The findings highlight the effectiveness of combining image processing with machine learning algorithms in agricultural research and crop improvement.

#### ACKNOWLEDGMENT

This research was supported by the National Institute of Food and Agriculture Department (NIFA) through the Evans Allen Grant Project, funded by the United States Department of Agriculture (USDA) (Grant no: GR015407/E/A # 20250/SI).

#### **Conflict of Interest**

The authors have declared that there are no competing interests.

## Authors Contribution

IS:Resources, Supervision, Conceptualization, Data curation, Writing review & editing. PI of the research project. RT: Data curation, Writing review & editing. IH: Writing, review & editing. MA: Data curation, Writing review.

## REFERENCES

Alam MK, Sams S, Rana ZH, Akhtaruzzaman M, Islam SN., 2020. Minerals, vitamin C, and effect of thermal processing on carotenoid composition in nine varieties of orange-fleshed sweet potato (*Ipomoea batatas* L.). Journal of Food Composition and Analysis, 92: 103582.

Amankwaah VA., 2012. Phenotypic and molecular characterization of released and elite sweetpotato varieties in Ghana compared with virus-tested putative ramets. 2012. https://api.semanticscholar.org/CorpusID:131211575.

Aywa AK, Nawiri MP, Nyambaka HN., 2013. Nutrient variation in colored varieties of Ipomoea batatas grown in Vihiga county, Western Kenya. International Food Research Journal, 20: 819–825.

Chen SP, Wang SY, Huang MY, Lin KH, Hua SM, Lu HH, Lai YC, Yang CM., 2018. Physiological and molecular analyses of chlorophyllase in sweet potatoes with different-colored leaves. South African Journal of Botany, 11: 272–279. https://doi.org/https://doi.org/10.1016/j.sajb.2017.11.021.

Chitwood D, Kumar R, Headland L, Ranjan A, Covington M, Ichihashi Y, Fulop D, Jiménez-Gómez J, Peng J, Maloof J, Sinha N., 2013. A quantitative genetic basis for leaf morphology in a set of precisely defined tomato introgression lines. Plant Cell, 25(7): 2465-2481. https://doi.org/10.1105/tpc.113.112391.

Du JX, Wang XF, Zhang GJ., 2007. Leaf shape based plant species recognition. AppliedMathematicsandComputation,185(2):883-893.https://doi.org/10.1016/j.amc.2006.07.072.

Gehan MA, Fahlgren N, Abbasi A, Berry JC, Callen ST, Chavez L, Doust AN, Feldman MJ, Gilbert KB, Hodge JG, Hoyer JS, Lin A, Liu S, Lizárraga C, Lorence A, Miller M, Platon E, Tessman M, Sax T., 2017. Plant CV v2: Image analysis software for high-throughput plant phenotyping. PeerJ., 5: e4088. https://doi.org/10.7717/peerj.4088.

Gunathilake KDPP, Ranaweera KKDS., 2016. Antioxidative properties of 34 green leafy vegetables. Journal of Functional Foods, 26: 176–186. https://doi.org/https://doi.org/10.1016/j.jff.2016.07.015.

Gupta S, Rosenthal DM, Stinchcombe J, Baucom RS., 2020. The remarkable morphological diversity of leaf shape in sweetpotato (*Ipomoea batatas*): the influence of genetics, environment, and GxE. New Phytologist, 225: 2183-2195. doi: 10.1111/nph.16286.

Huang X, Tu Z, Xiao H, Li Z, Zhang Q, Wang H, Hu Y, Zhang L., 2013. Dynamic highpressure micro fluidization-assisted extraction and antioxidant activities of sweet potato (Ipomoea batatas L.) leaves flavonoid. Food and Bioproducts Processing, 91: 1– 6. https://doi.org/https://doi.org/10.1016/j.fbp.2012.07.006.

Islam MH, Kondo N, Ogawa Y, Fujiura T, Suzuki T, Fujitani S., 2017. Detection of infertile eggs using visible transmission spectroscopy combined with multivariate analysis. Engineering in Agriculture, Environment and Food, 10: 115–120. https://doi.org/https://doi.org/10.1016/j.eaef.2016.12.002

Islam MH, Rahman A, Rana MS., 2021. Potato grading based on size features by machine vision technique. Journal of the Bangladesh Agricultural University, 19(4): 528–532. Available online: https://doi.org/10.5455/JBAU.123862.

Islam S, Adam Z, Ishrar I., 2020. Comparative study on some in vitro biological activities of freeze-dried leaf extracts of six advanced accessions of Ipomoea batatas (L.) Lam. Nutrition & Food Science, 10: 1-7. https://juniperpublishers.com/nfsij/NFSIJ.MS.ID.555776.php

Islam S, Chowdhury A, Akanda J., 2023. Mineral, vitamin C, and oxalate contents in the leaves of 26 selected sweetpotato (*Ipomoea batatas* L.) accessions growing in the southern United States. Journal of Agricultural Environmental and Consumer Sciences, 23: 49-54.

Islam S, Jalaluddin M, Garner J, Yoshimoto M, Yamakawa O., 2005. Artificial shading and temperature influenced on anthocyanin composition of Ipomoea batatas leaves. HortScience, 176-180. https://doi.org/10.21273/HORTSCI.40.1.176

Islam S., 2006. Sweetpotato (*Ipomoea batatas* L.) leaf: Its potential effect on human health and nutrition. Journal of Food Science, 71: 13–21. https://doi.org/https://doi.org/10.1111/j.1365-2621.2006.tb08912.x.

Islam S., 2009. Polyphenol contents and caffeic acid derivatives from leaves of Ipomoea batatas genotypes. Acta Horticulturare, 841, 527-530. https://doi.org/10.17660/ActaHortic.2009.841.75

Islam S., 2014. Medicinal and Nutritional Qualities of Sweetpotato Tips and Leave. Published by Cooperative Extension Service, FSA 6135: 1-4.

Islam S., 2016. Some bioactive constituents, antioxidant, and antimutagenic activities in the Leaves of *Ipomoea batatas* Lam. Genotypes. American Journal of Food Science & Technology, 4(3): 70-80. https://pubs.sciepub.com/ajfst/4/3/3/

Islam S., 2024. Sweetpotatoes [*Ipomoea batatas* (L.) lam]: the super food of the Next Century? An intensive review on their potential as a sustainable and versatile food source for future generations. *CyTA* - Journal of Food, 22(1). https://doi.org/10.1080/19476337.2024.2397553

Islam, S. 2008. Potential chemo-preventative properties isolated from Ipomoea batatas leaves, in Functional Foods and Chronic Diseases. Functional Food Center at D & A Inc., TX, USA. 2008, 96-109.

Islam, S. 2019. Antimutagenicity of the water extracts, radical scavenging activity, and phenolic acids in the tops of diverse *Ipomoea batatas* (L.) Lam. Advances in Biomedical Sciences, 4: 46-51.

Jackson DM, Harrison HF, Jarret RL, Wadi PA., 2020. Phhenotypic variation in leaf morphology of the USDA, ARS sweetpotato (*Ipomoea batatas*) germplasm collection. HortScience, 55(4):465–475. https://doi.org/10.21273/HORTSCI14703-19

Jia R, Tang C, Chen J, Zhang X, Wang Z., 2022. Total phenolics and anthocyanins contents and antioxidant activity in four different aerial parts of leafy sweet potato (*Ipomoea batatas* L.). Molecules, 2022: 27. https://doi.org/10.3390/molecules27103117.

Kumar J, Pratap A, Kumar S., 2015. Plant Phenomics: An Overview, in Kumar, J., Pratap, A., Kumar, S. (Eds.), Phenomics in Crop Plants: Trends, Options and Limitations. Springer India, New Delhi. 2015, 1–10. https://doi.org/10.1007/978-81-322-2226-2\_1.

Lako J, Trenerry VC, Wahlqvist M, Wattanapenpaiboon N, Sotheeswaran S, Premier R., 2007. Phytochemical flavonols, carotenoids, and the antioxidant properties of a wide selection of Fijian fruits, vegetables, and other readily available foods. Food Chemistry, 101: 1727-1741. https://doi.org/https://doi.org/10.1016/j.foodchem.2006.01.031.

Li M, Jang GY, Lee SH, Kim MY, Hwang SG, Sin HM, Kim HS, Lee J, Jeong HS., 2017. Comparison of functional components in various sweet potato leaves and stalks. Food Science and Biotechnology, 28;26(1): 97-103. doi: 10.1007/s10068-017-0013-6. PMID: 30263515; PMCID: PMC6049489.

Liang J, Doermann D, Li H., 2005. Camera-Based analysis of text and documents: a survey. International Journal on Document Analysis and Recognition, 7(2-3): 84-104. https://doi.org/10.1007/s10032-004-0138-z.

Lu Y, Wang J, Fu L, Yu L, Liu Q., 2023. High-throughput and separating-free phenotyping method for on-panicle rice grains based on deep learning. Frontiers in Plant Science, 14:1219584. doi: 10.3389/fpls.2023.1219584.

Mahmood N, Moore PS, Tommasi ND, Simone FD, Colman S, Hay AJ, Pizza C., 1993. Inhibition of HIV infection by caffeoylquinic acid derivatives. Antiviral Chemistry and Chemotherapy, 4: 235-240. https://doi.org/10.1177/095632029300400406

Matsui T, Ebuchi S, Fujise T, Abesundara KJM, Doi S, Yamada H, Matsumoto K., 2004. Strong antihyperglycemic effects of the water-soluble fraction of Brazilian propolis and its bioactive constituent, 3,4,5-tri-O-caffeoylquinic acid. Biological and Pharmaceutical Bulletin, 27: 1797-1803. https://pubmed.ncbi.nlm.nih.gov/15516726/

Matsui T, Ebuchi S, Kobayashi M, Fukui K, Sugita K, Terahara N, Matsumoto K., 2002. The Antihyperglycemic effect of diacylated anthocyanin derived from *Ipomoea batatas* cultivar Ayamurasaki can be achieved through the  $\alpha$ -glucosidase inhibitory action. Journal of Agricultural and Food Chemistry, 50: 7244-7248. https://pubmed.ncbi.nlm.nih.gov/12452639/

Wang M, Fu B, Fan J, Wang Y, Zhang L, Xia C., 2023. Sweet potato leaf detection in a natural scene based on faster R-CNN with a visual attention mechanism and DIoU-NMS. Ecological Informatics, 73, 101931. https://doi.org/10.1016/j.ecoinf.2022.101931

Ngailo S, Shimelis H, Sibiya J, Mtunda K., 2013. Sweet potato breeding for resistance to sweet potato virus disease and improved yield progresses and challenges. African Journal of Agricultural Research, 8: 3202–3215. https://doi.org/10.5897/AJAR12.1991.

Ogunlesi M, Okiei W, Azeez L, Obakachi V, Osunsanmi M, Nkenchor G., 2010. Vitamin C contents of tropical vegetables and foods are determined by voltammetric and titrimetric methods and their relevance to the medicinal uses of the plants. International Journal of Electrochemical Science, 5: 105–115.

Onyango CM, Shibairo SI, Imungi JK, Harbinson J., 2008. The physical and chemical characteristics and some nutritional value of the vegetable Amaranth sold in Nairobi - Kenya. Ecology of Food and Nutrition, 47(4): 382-398.

Peluso G, Feo VD, Simone FD, Bresciano E, Vuotto ML., 1995. Studies on the inhibitory effects of caffeoylquinic acids on monocyte migration and superoxide anion production. Journal of Natural Products, 58: 639-646. https://doi.org/10.1021/np50119a001

Radek M, Savage GP., 2008. Oxalates in some Indian green leafy vegetables. International Journal of Food Sciences and Nutrition, 59(3): 246-260.

Rosero A, Granda L, Pérez JL, Rosero D, Burgos-Paz W, Martı'nez M, Morelo J, Iva'n Pastrana I, Burbano E, Morales, A., 2019. Morphometric and colorimetric tools to dissect morphological diversity: an application in sweet potato [Ipomoea batatas (L.) Lam.]. Genetic Resources and Crop Evolution, 66: 1257-1278. https://doi.org/10.1007/s10722-019-00781-x

Rumbaoa RGO, Cornago DF, Geronimo IM., 2009. Phenolic content, and antioxidant capacity of Philippine sweet potato (*Ipomoea batatas*) varieties. Food Chemistry, 113: 1133–1138. https://doi.org/https://doi.org/10.1016/j.foodchem.2008.08.088.

Shimozono H, Kobori M, Shinmoto H, Tsushida T., 1996. Suppression of the melanogenesis of mouse melanoma B 16 cells by sweetpotato extract. Nippon Shokuhin Kagaku Kogaku Kaishi, 43: 313-317. https://www.semanticscholar.org/paper/Suppression-of-the-Melanogenesis-of-

Mouse-Melanoma-Shimozono-Kobori/c91c89de4eb09bb6c70461d8ddd93d37a8ae17e5

Si Y, Sankaran S, Knowles N, Bgood J., 2016. Potato tuber length-width ratio assessment using image analysis. American Journal of Potato Research, 94: 1–6. https://doi.org/10.1007/s12230-016-9545-1.

Smith TP, Stoddard S, Shankle M, Schultheis J., 2009. Sweetpotato Production in the United States. In: Loebenstein G, Thottappilly G, editors. The Sweetpotato, Dordrecht. Springer Netherlands, 287–323. https://doi.org/10.1007/978-1-4020-9475-0\_14.

Statista., 2016. Per capita consumption of fresh sweet potatoes in the United States from 2000 to 2014. 12 Sept. 2016. http://www.statista.com/statistics/257307/per-capita-consumption-of-fresh-sweet-potatoes-in-the-us/

Su WH, Xue H., 2021. Imaging spectroscopy and machine learning for intelligent determination of potato and sweetpotato quality. Foods, 10(9): 2146. https://doi.org/10.3390/foods10092146

Suda I, Yamakawa O, Matsugano K, Sugita K, Asuma K, Irisa K, Tokumaru F., 1998. Changes of serum  $\gamma$ –GTP, GOT, and GPT levels in hepatic function-weak in subjects by ingestion of high anthocyanin sweetpotato juice. Nippon Shokuhin Kagaku Kogaku Kaishi, 45: 611–617.

Sun G, Chen X, Ren J, Zhang A, Jia X., 2017. Stratified spectral mixture analysis of medium-resolution imagery for impervious surface mapping. International Journal of Applied Earth Observation and Geoinformation, 60: 38-48. https://doi.org/10.1016/j.jag.2017.04.006.

Taira J, Taira K, Ohmine W, Nagata J., 2013. Mineral determination and anti-LDLoxidation activity of sweet potato (*Ipomoea batatas* L.) leaves. Journal of FoodCompositionandAnalysis,29:117-125.https://doi.org/https://doi.org/10.1016/j.jfca.2012.10.007.

Toeller M., 1994. alpha- Glucosidase inhibitors in diabetes: efficacy in NIDDM subjects. European Journal of clinical Investigation, 24: 32-35. https://doi.org/10.1111/j.1365-2362.1994.tb02253.x

Wang M, Fu B, Fan J, Wang Y, Zhang L, Xia C., 2023. Sweet potato leaf detection in a natural scene based on faster R-CNN with a visual attention mechanism and DIoU-NMS. Ecological Informatics, 73: 101931, 2023.

Wang S, Nie S, Zhu F., 2016. Chemical constituents and health effects of sweet potato.FoodResearchInternational,89:90-116.https://doi.org/https://doi.org/10.1016/j.foodres.2016.08.032.

Xu W, Liu L, Hu B, Sun Y, Ye H, Ma D, Zeng X., 2010. TPC in the leaves of 116 sweetpotato (*Ipomoea batatas* L.) varieties and Pushu 53 leaf extracts. Journal of FoodCompositionandAnalysis,23:599-604.

https://doi.org/https://doi.org/10.1016/j.jfca.2009.12.008.

Yamakawa O, Yoshimoto M., 2002. Sweetpotato is a new food material with physiological functions. In 'Proceedings 1st International Conference on Sweetpotato, Food and Health for the Future. Acta Horticulture, 583: 179–185. https://www.actahort.org/books/583/583\_20.htm

Yao Q,, Guan Z, Zhou Y, Tang J, Hu Y, Yang B., 2009. Application of support vector machine for detecting rice diseases using shape and color texture features. International Conference on Engineering Computation, 79–83. https://doi.org/10.1109/ICEC.2009.73.

Yoshimoto M., 2001. New trends of processing and use of sweetpotato in Japan. Farming Japan, 35: 22-28.

Zhang D, Cervantes J, Huamán Z, Carey E, Ghislain M., 2000. Assessing genetic diversity of sweet potato (*Ipomoea batatas*) cultivars from tropical America using AFLP. Genetic Resources and Crop Evolution, 47(6): 659–665. https://doi.org/10.1023/A:1026520507223.

Zhao C, Zhang Y, Du JJ, Guo X, Wen W, Gu S, Wang J, Fa J., 2019. Crop Phenomics: Current Status and Perspectives. Frontiers in Plant Science, 10. https://doi.org/10.3389/fpls.2019.00714.